# CVMPROAc001 p003 SARSMpro in pNIC transformation, expression and purification

PAGE24-00585

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**Experiment Started:** 

Projects: Expression; Purification; ASAP Related Pages: PAGE23-00603; PAGE23-00782

Referenced by:

Tags:

#### **Transformation**

Plasmid was purchased from Twist and dissolved in 10uL of nuclease-free water. Transformation carried out per the standard protocol.

CVMPROA-c001

#### **Expression**

22/03/2024

6x1L Invitrogen TB + Kan + 4mL glycerol after autoclave + Kan50 1mL of ON from transformation was used to inoculate per L of TB. Grown at 37oC 200rpm until OD~1.8, induced with 0.5mM IPTG Temperature reduced to 18oC and speed to 180rpm. Grown overnight.

Harvest next day. Total pellet weight = 88g

## IMAC, cleavage and rIMAC

MGSSHHHHHHGSDSEVNQEAKPEVKPEVKPETHINLKVSDGSSEIFFKIKKTTPLRRLMEAFAKRQGKEMDSLRFLYDGIRIQA DQTPEDLDMEDNDIIEAHREQIGGSGFRKMAFPSGKVEGCMVQVTCGTTTLNGLWLDDVVYCPRHVICTSEDMLNPNYEDLL IRKSNHNFLVQAGNVQLRVIGHSMQNCVLKLKVDTANPKTPKYKFVRIQPGQTFSVLACYNGSPSGVYQCAMRPNFTIKGSFL NGSCGSVGFNIDYDCVSFCYMHHMELPTGVHAGTDLEGNFYGPFVDRQTAQAAGTDTTITVNVLAWLYAAVINGDRWFLNR FTTTLNDFNLVAMKYNYEPLTQDHVDILGPLSAQTGIAVLDMCASLKELLQNGMNGRTILGSALLEDEFTPFDVVRQCSGVTFQ

Purification ID: CVMPROA-p001 (labelled p003 on bag)

#### **Buffers:**

Lysis buffer - 50 mM HEPES pH 7.4, 150 mM NaCl, 5% glycerol, 0.5mM TCEP

Wash buffer - 50 mM HEPES pH 7.4, 150 mM NaCl, 5% glycerol, 0.5mM TCEP Elution Buffer - 50 mM HEPES pH 7.4, 150 mM NaCl, 5% glycerol, 0.5mM TCEP, 500mM imidazole Gel Filtration Buffer (SEC) - 50 mM HEPES pH 7.4, 150 mM NaCl, 5% glycerol, 0.5mM TCEP

# no imidazole in binding and wash step due to previous difficulties with target not binding to IMAC resin

- 1. Lysis buffer supplement with 1:4000 dilution of benzonase, 0.5 mg/mL lysozyme. Incubate for 30min at RT.
- 2. Sonicated on ice at 50% amplitude for a total of 7-minute sonication time (4 seconds on 12 seconds off) with thick probe.
- 3. Clarified lysate by centrifugation at 18,000rpm, 4°C for 1 hour. Used JLA16.250 rotor. Supernatant poured into clean beaker.

### **IMAC**

- 1. Wash and equlibrate 5mL bed volume of Ni Sepharose resin on gravity flow column, first with distilled water, then with wash buffer.
- 2. Resuspend resin with lysis buffer and add to beaker containing clarified supernatant.
- 3. Stir gently in cold room for 30mins. Pour only gravity flow column and allow to flow though.
- 4. Wash resin with 25mL wash buffer twice.
- 5. Elute with 7.5mL elution buffer, 2 elutions carried out

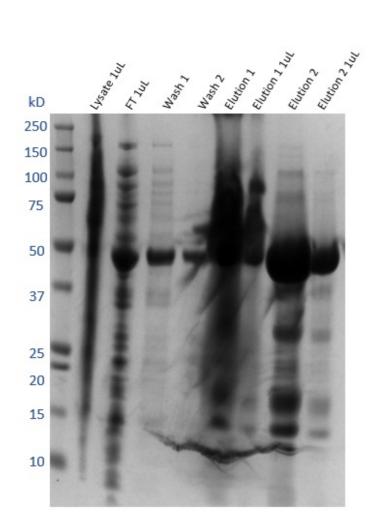
## **TEV** cleavage

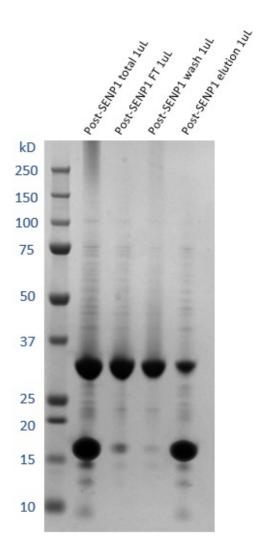
- 1. Desalt elution with HiLoad 10/26 desalting column on ATKA. Desalt into lysis buffer.
- 2. Pool desalted protein from the fractions
- 3. Add SENP1 in 1:300 OD ratio. Leave to incubate in cold room.

### rIMAC and Gel filtration

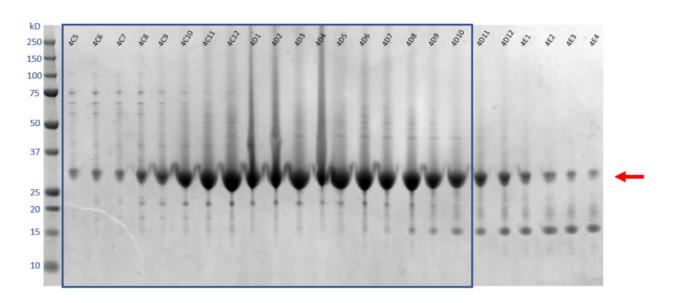
- 1. in morning, IMAC resin washed with some lysis buffer to remove imidazole.
- 2. cleavage mix passed through the resin 1 time (flow rate was a bit slow this time
- 3. Wash resin with 10mL wash buffer
- 4. Elute to see what stuck to the resin
- 5. rIMAC FT concentrated in 10kDa MWCO yellow vivaspin concentrators to final volume of ~5mL
- 6. Injected onto Superdex s200 16/60 column and run in SEC buffer at 1mL/min
- 7. After SEC, peaks were analysed by SDS-PAGE

**IMAC** results





# **SEC result**



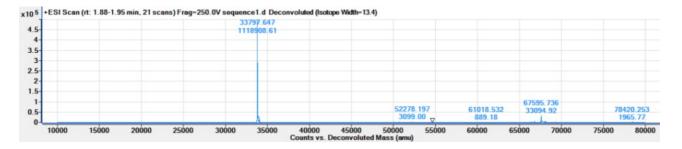
## **Final sample**

Fraction 4C5 - 4D10 were pooled and concentrated in the yellow vivaspin 10kDa MWCO concentrators

Final concentration: 26.96 mg/mL, 47\*100 + 50 uL

Total yield: 128.1 mg

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## final sample

