

SRF-Bio-ASAP XX01ZVNS2B-c001 expression and purification

PAGE23-01623

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Date Started: **2023-Oct-02**

Experiment Started:

Projects: **Expression;Purification;ASAP**

Related Pages: **PAGE22-01775;PAGE23-00292;PAGE23-01401**

Referenced by:

Tags: **ASAP**

Info

XX01ZVNS2B-c001

MHHHHHHSSMSPILGYWKIKGLVQPTRLLEYLEEKYEEHLYERDEGDKWRNKKFELGLEFPNLPYYIDGDVKLTQSMAIIRYI
ADKHNMLGGCPKERAIEISMLEGAVLDIRYGVSR IAYSKDFETLKVDFLSKLPEMLKMFEDRLCHKTYLNGDHVTHPDFMLYDA
LDVVLYMDPMCLDAFPKLVCFKKRIEAI PQIDKYLKSSKYIAWPLQGWQATFGGGDHPPKSGGGSENLYFQ//SMGKSVD MYIE
RAGDITWEKDAEVTGNSPRLDVALDESGD FSLVEE

WITH TAG:

MW:32812.54

E(red): 51340

PI: 5.47

WITHOUT TAG

NS2B

SMGKSVD MYIERAGDITWEKDAEVTGNSPRLDVALDESGD FSLVEE

MW: 5063.51

E(red): 6990

PI: 3.98

NS3

MAPKEVKKGETT DGVYRVMTRRL LGSTQVG VGMQEGVFHTMWHVTKGAALRS GEGRLDPYWG DVKQDLVSYCGPWKLDAAWDGLSEVQL
LAVPPGERAKNIQTLP G I FKT KDGDIGA VALDYPAGTSGSPILDKCGRVIGLYGNGVVIKNGSYVSAITQGKREEETPVE

MW: 18095.66

E(red) 30940

NS2B+NS3

MW: 23309.34

E(red): 37930

expect to see three bands after cleavage and rIMAC: ~25kDa, ~18kDa and ~12kDa

Because of coexpression

Previously grown in regular TB, induced at ~OD₂ with 5mM IPTG (that's a lot), 18oC overnight expression

33g total pellet wet weight from 2L culture

Purified in standard HEPES buffer. (cleave in 20mM imidazole)

Final buffer: 50 mM HEPES, pH 7.5, 500 mM NaCl, 20 mM Imidazole, pH 7.0, 5% glycerol, 1 mM TCEP

Clone 6 GST in Mike's page

Expression ID: XX01ZVNS2B-e004

Purification ID: XX01ZVNS2B-p004

Expression

2 Oct 2023

10mL ON culture:

RR3 Glycerol stock from ASAP-1 H06 inoculated in 10mL of 2xLB, grown 37°C 220rpm overnight

3 Oct 2023

Add 1mL of ON culture into 100mL 2xLB+Kan, grow 37°C 220rpm overnight

4 Oct 2023

Add 10mL each into 2x1L TB media (16mL 100% glycerol added after autoclave) + Kan

Grow at 37°C, 250rpm until OD~2

Induce expression with 5mM IPTG, 18°C, 200rpm overnight.

Harvested next day. split into 2x3L: 48 and 51g.

Purification

Buffers:

Base buffer - 50 mM HEPES pH7.5, 500 mM NaCl, 5% glycerol, **1mM TCEP**

Wash buffer 1 - 50 mM HEPES pH 7.5, 500 mM NaCl, 5% glycerol, **1mM TCEP**, 30mM imidazole

Wash buffer 2 - 50 mM HEPES pH 7.5, 500 mM NaCl, 5% glycerol, **1mM TCEP**, 50mM imidazole

Elution Buffer - 50 mM HEPES pH 7.5, 500 mM NaCl, 5% glycerol, **1mM TCEP**, 500mM imidazole

TEV cleavage buffer - 50 mM HEPES pH 7.5, 500 mM NaCl, 5% glycerol, **1mM TCEP**, 20mM imidazole

Gel Filtration Buffer (SEC) - 50 mM HEPES pH 7.5, 500 mM NaCl, 5% glycerol, **1mM TCEP**, 20mM imidazole

2L pellet thawed and homogenised with lysis buffer to 250mL, supplemented with 1% TX-100, 0.5mg/mL HEWL, 1:4000 benzonase, 20mM imidazole.

Sonicate 40% 10 min 2s on 4s off over ice

Pour clarified supernatant over 3mL Ni Sepharose 6 FF

Wash 50mL 30imi, 50mL 50imi, elute 2x7.5mL 500imi

(used 1OD=1mg/mL)

E1: 18.28 mg/mL

E2: 13.02 mg/mL

after desalt: 9.16 mg/mL, 13mL

STICKY PROTEIN?

ACCIDENTALLY ADDED SENP1 INSTEAD OF TEV

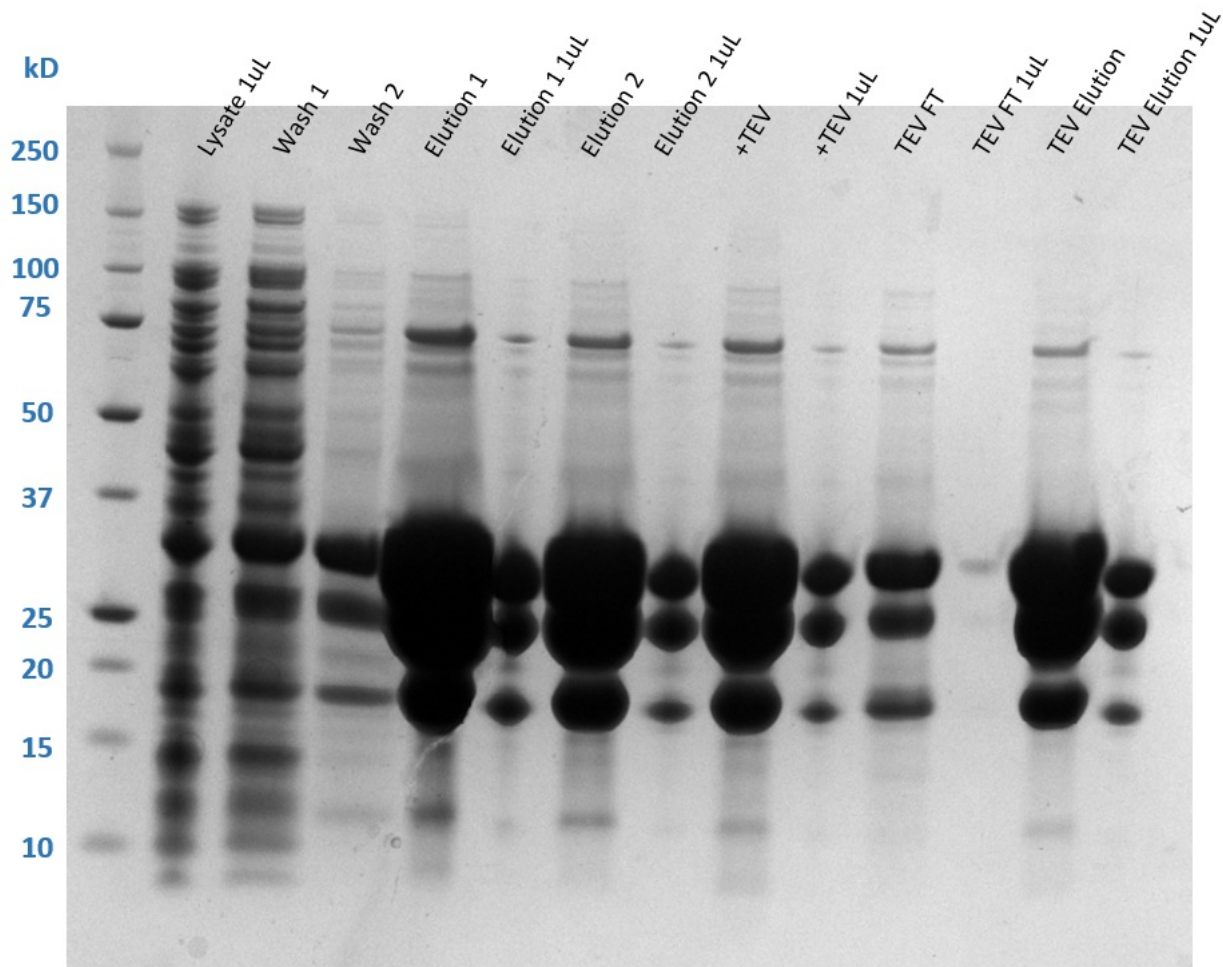
poured over rinsed IMAC resin 2 times, FT collection. Elution with 5mL elution buffer collected.

FT concentrated to ~3mL with 10kDa amicon concentrators and loaded onto s75 column.

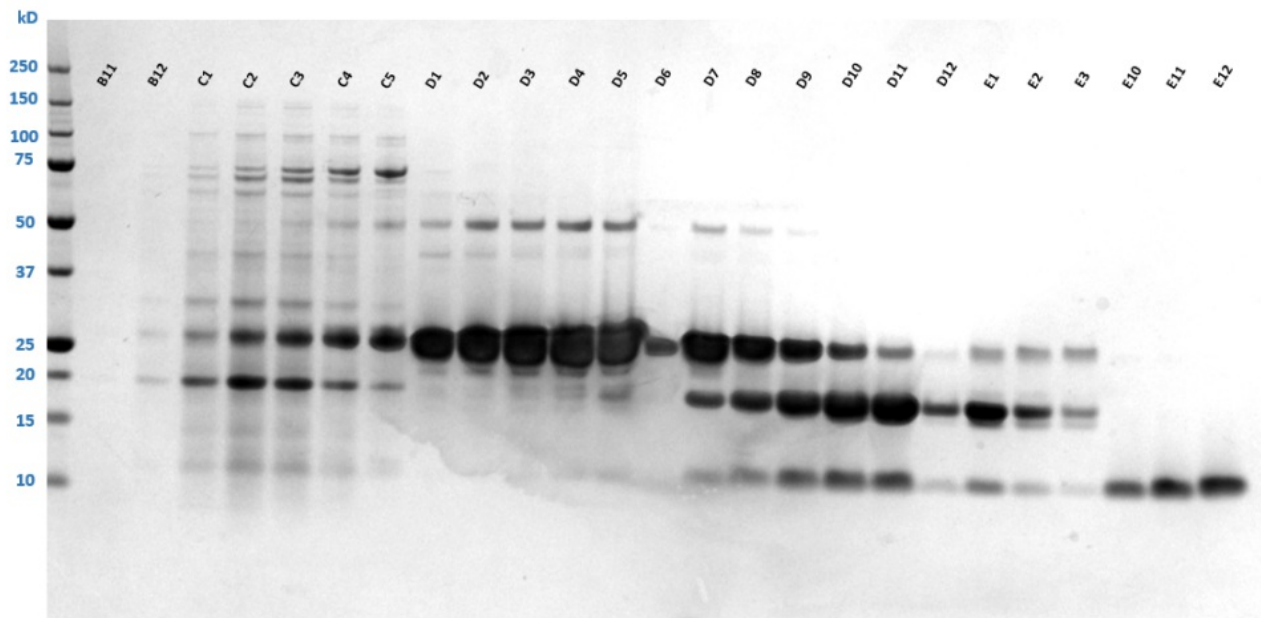
To rescue: pool all protein- containing fractions from s75
rIMAC elution (5mL) diluted to 50mL to reduce imidazole concentration to 50mM. Add to the s75 fractions.
TEV added 1:100. Incubated overnight in cold room.

the next day: concentrated to ~3mL and injected to s75 again

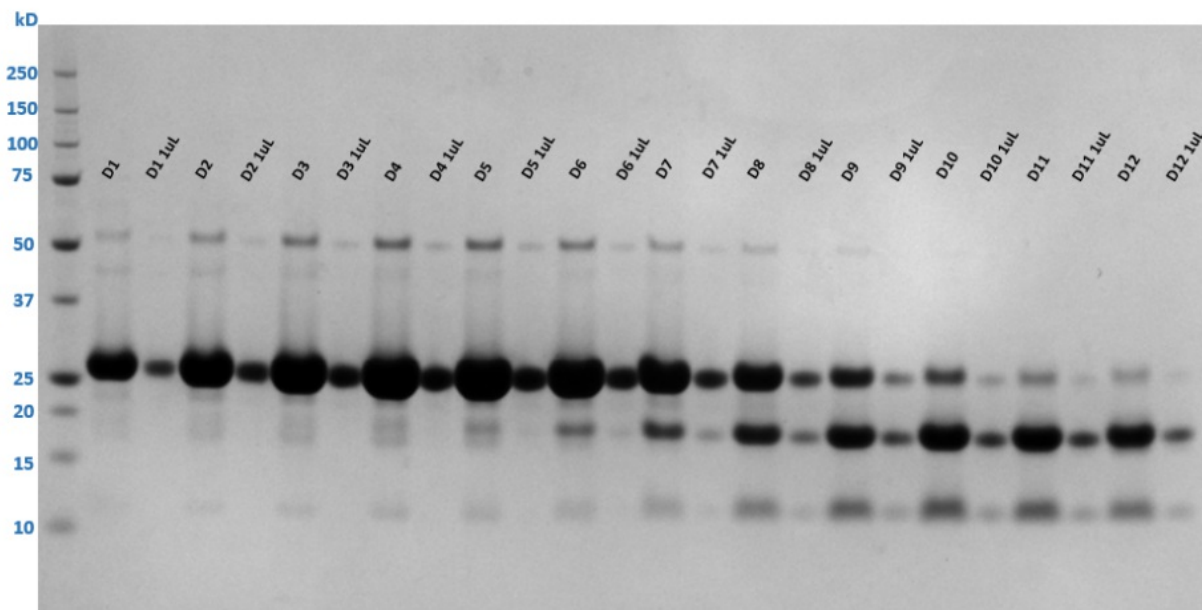
IMAC result



Second round TEV - SEC result



SEC rerun with more DTT and longer boiling



Batch 2

Second 3L also processed with following modification:

- rIMAC is carried out by pouring cleavage mix over the used resin (washed) 3x

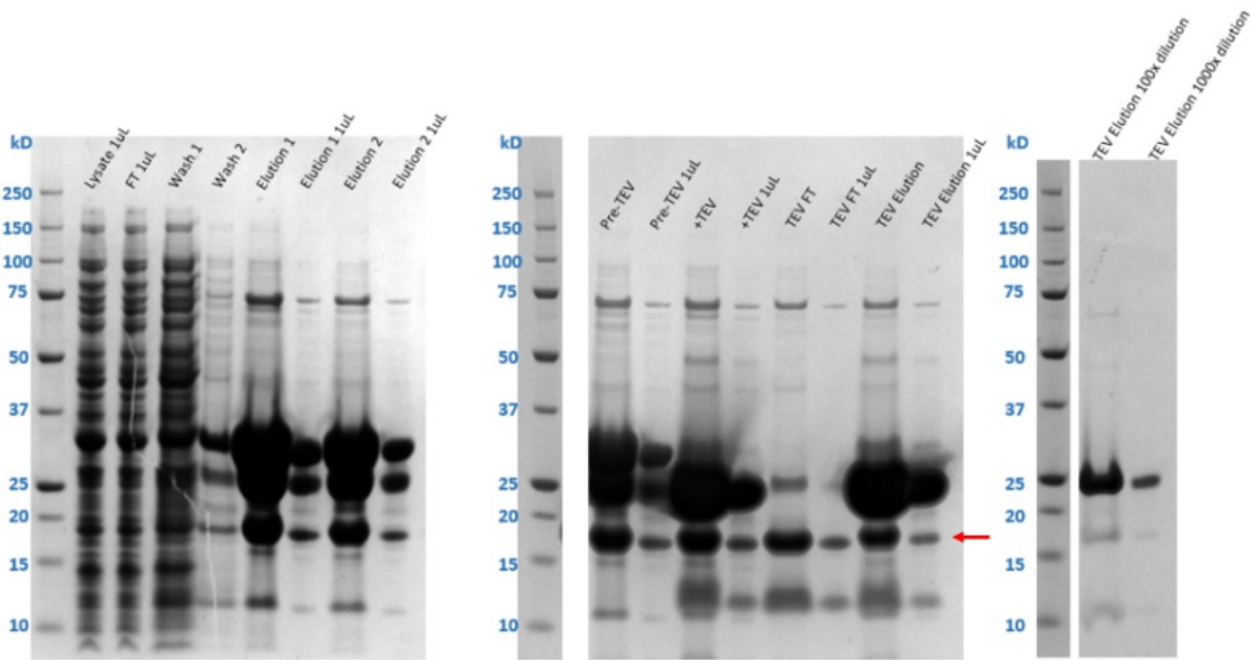
This helped clean up the FT from TEV and uncleaved products significantly.

The protein seems to stick to the resin?

A280 decreased by almost 1/5 this time as well

Bands corresponding to cleaved protein also appear in TEV elution (500imi) and is quite dark.

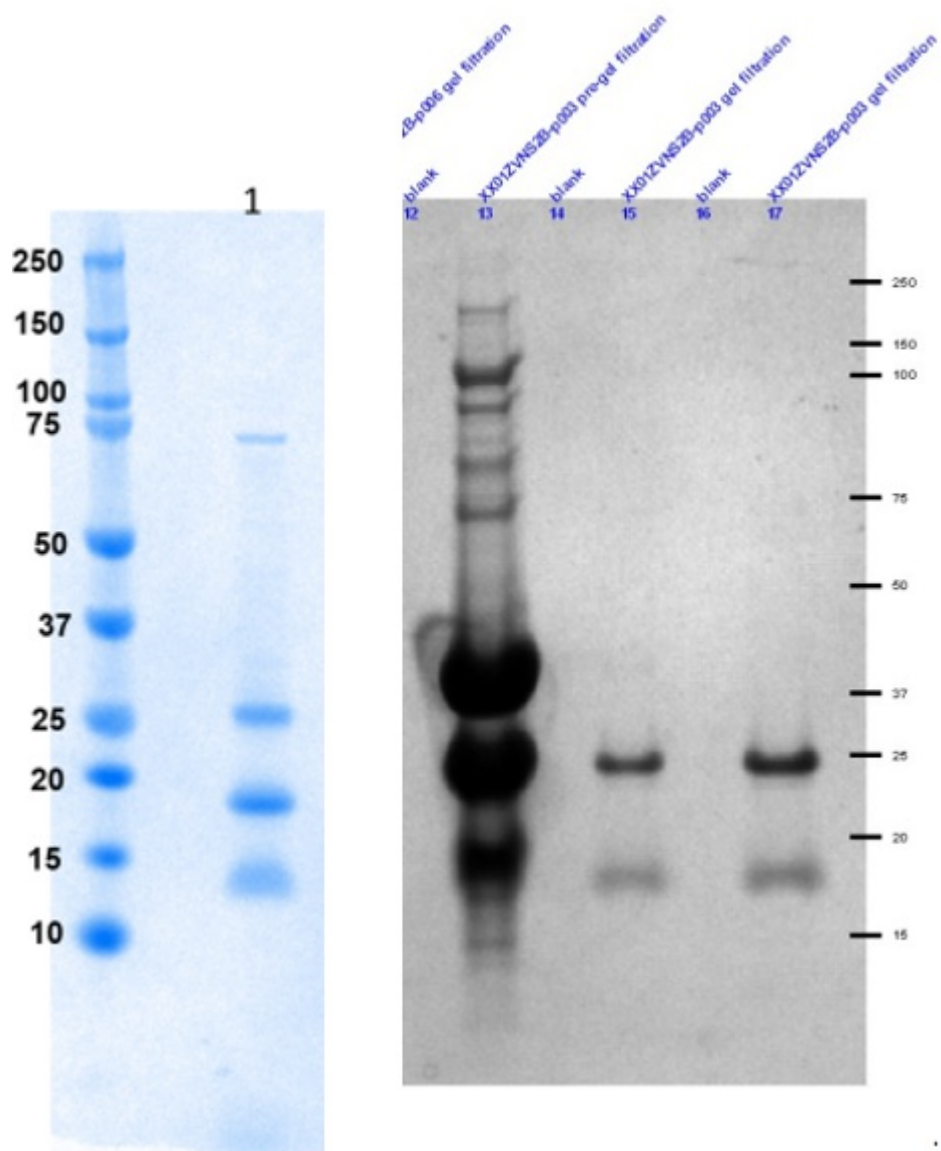
Batch 2 IMAC and cleavage results



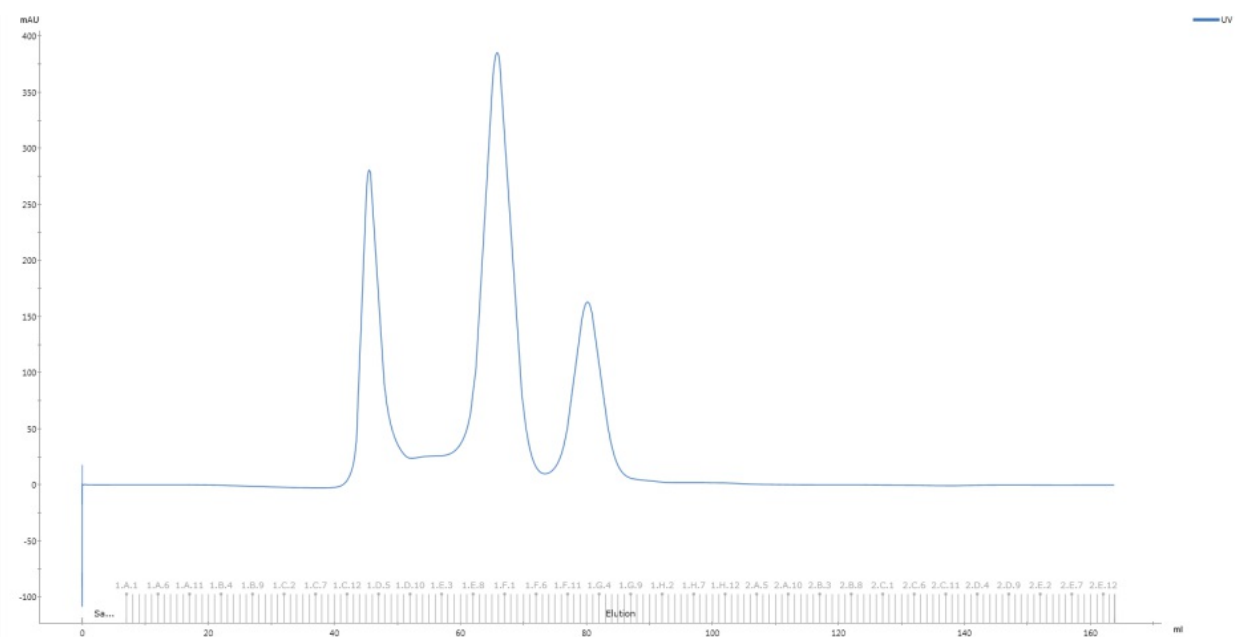
Batch 2 IMAC and cleavage results arrow

Red arrow = cleaved NS3

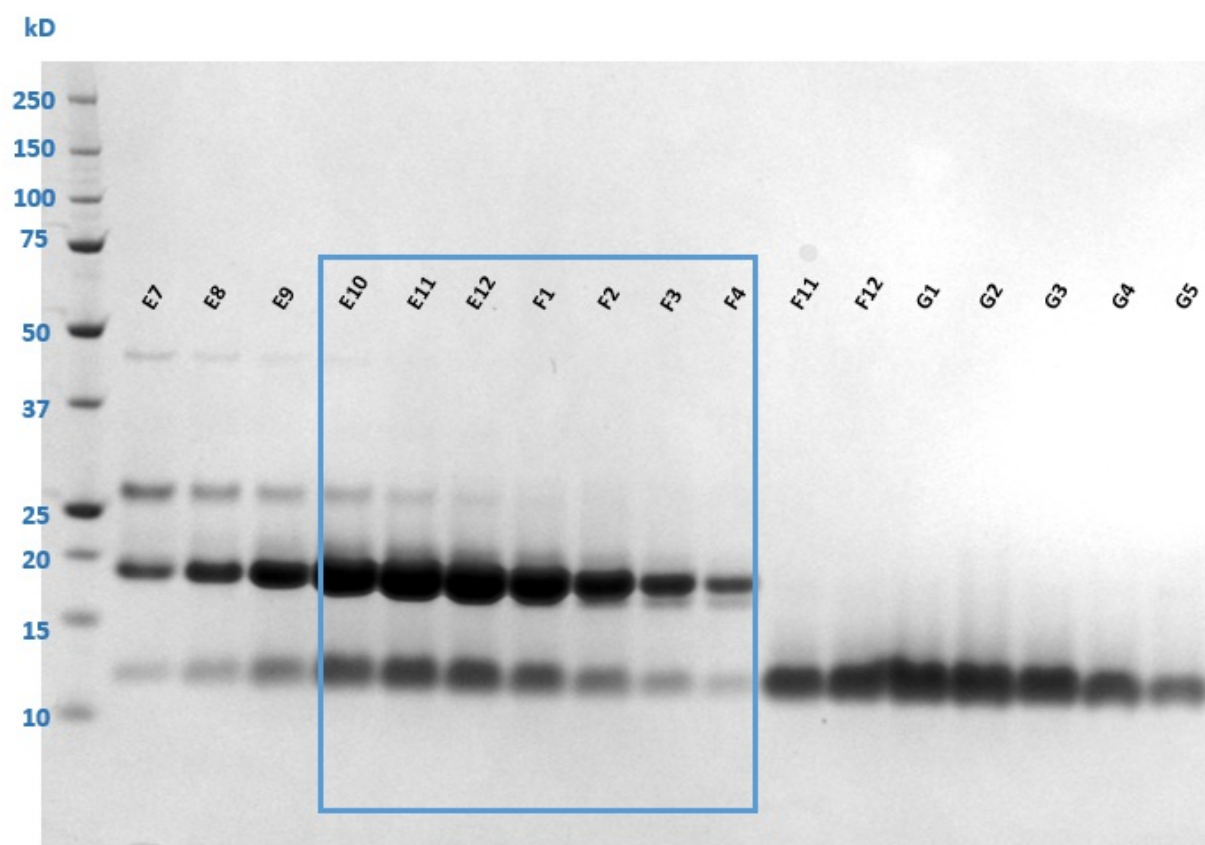
Comparison with Mike and Mel's previous purifications



Chromatogram



Batch 2 SEC



Concentration and final sample

Fraction D12-E1 and fraction E10-F4 from SEC 2 were pooled and concentrated in 10kDa MWCO amicon concentrators, then Vivaspin 500 10kDa concentrators.

Final concentration is 42.25 mg/mL, 10x10uL aliquots.

Yield = 4.23 mg