TH DAB staining

- 1. Wash a full set of sections (6-8 sections) in a well in PBS, $5 \text{ min} \times 3$ (if stored in cryoprotective solution).
- 2. Incubate sections in 1% H₂O₂ (1 ml 30% H₂O₂ in 29 ml TBST), 30 min at RT.
- 3. Wash sections in TBST 5 min \times 3.
- 4. Block sections in 5% NGS/TBST, 1 hour at RT.
- 5. Incubate sections in rabbit anti-TH (#SA-497), 1:3000 in TBST, 4C overnight.
- 6. Wash sections in TBST, $10 \min \times 3$.
- 7. Incubate sections in anti-rabbit biotinylated (#OS03B), **1:3000**, 1hour at RT.
- 8. Prepare ABC (Avidin-Biotin-Complex, Vectastain ABC Kit, # PK-6100): 1 drop A and 1 drop B in 2.5 ml TBST, at least 30 min before use.
- 9. Wash sections in TBST, $10 \min \times 3$.
- 10. Incubate sections in ABC, 30 min at RT.
- 11. Prepare DAB solution (Sigma FAST, DAB Peroxidase Substrate Tablet Set), 1 DAB and 1 Urea Hydogen Peroxide tablet in 15 ml distilled water.
- 12. Wash sections in TBST, $10 \text{ min} \times 3$.
- 13. Incubate sections in 1 ml DAB, 1-2 min.
- 14. Wash sections in TBST, $5 \min \times 3$.
- 15. Mount the sections on positive charged plus slides.