

## NEBNext<sup>®</sup> Multiplex Oligos for Illumina<sup>®</sup> (Unique Dual Index UMI Adaptors DNA Set 1)

NEB #E7395S/L

96/384 reactions

Version 1.0\_3/20

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### The NEBNext Multiplex Oligos for Illumina (Unique Dual Index UMI Adaptors DNA Set 1) Includes

*The volumes provided are sufficient for preparation of up to 96 reactions (NEB #E7395S) and 384 reactions (NEB #E7395L).  
 All reagents should be stored at –20°C.*

NEBNext Primer Mix

NEBNext UMI Adaptor Dilution Buffer

NEBNext UMI DNA Adaptor Plate

Each well contains a unique dual index UMI adaptor (S size contains 1 plate, L size contains 4 plates)

*For the list of additional materials required, please check the manual for your NEBNext Library Prep Kit.*

### Overview

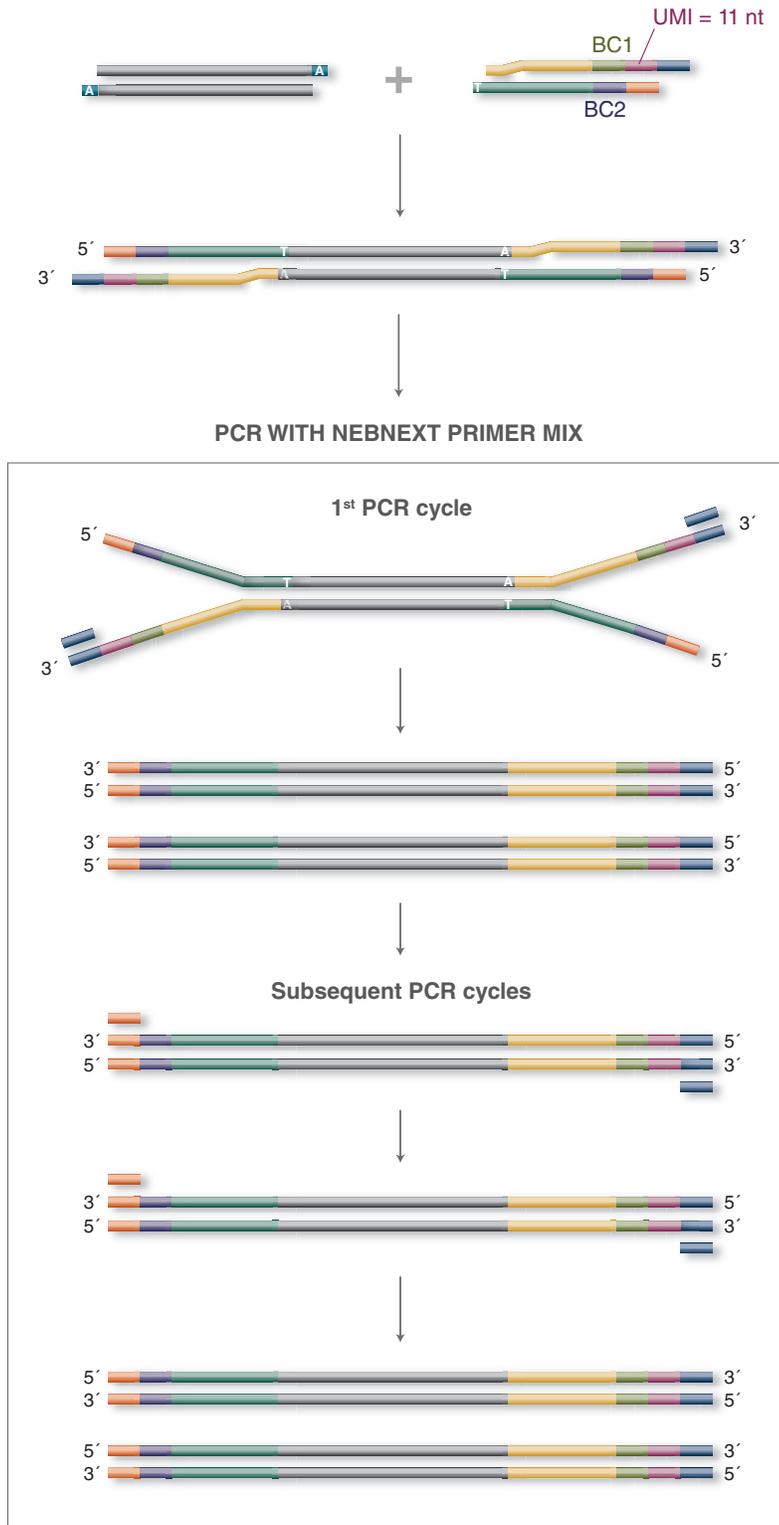
The NEBNext Multiplex Oligos for Illumina (Unique Dual Index UMI Adaptors DNA Set 1) contains adaptors and primers that are ideally suited for multiplex sample preparation for next-generation sequencing on the Illumina platform (Illumina, Inc.). Each kit component must pass rigorous quality control standards, and for each new lot the entire set of reagents is functionally validated together by construction and sequencing of indexed libraries on an Illumina sequencing platform.

For larger volume requirements, customized and bulk packaging is available by purchasing through the OEM/Bulks department at NEB. Please contact [OEM@neb.com](mailto:OEM@neb.com) for further information.

## Workflow

Designed for use in library prep for DNA, ChIP DNA and RNA (but not Small RNA), the NEBNext Unique Dual Index UMI (Unique Molecular Identifier) Adaptors enable high-efficiency adaptor ligation and high library yields. These adaptors contain all necessary sequences for sequencing on the Illumina platform, and are compatible with PCR-free applications and sample pooling prior to PCR amplification. The incorporation of a 11 base UMI allows 1) accurate identification and removal of duplicate reads, and 2) consensus sequence building and error correction, ideally suited for accurate analysis of quantitative NGS data. The 96 8-base unique dual index UMI adaptors included in this kit are packaged in a single-use 96-well plate with a pierceable foil seal. NEBNext Oligos can be used with NEBNext products, and with other standard Illumina-compatible library preparation protocols that are based on TA single base overhang ligation.

**Figure 1. Workflow demonstrating the use of NEBNext Multiplex Oligos for Illumina (Unique Dual Index UMI Adaptors DNA Set 1).**



## **Library Preparation Kits for use with NEBNext Unique Dual Index UMI Adaptors DNA Set 1**

Please refer to the kit specific protocol. The following kits are designed for use with the NEBNext Multiplex Oligos for Illumina:

- #E7805, NEBNext Ultra™ II FS DNA Library Prep Kit for Illumina
- #E6177, NEBNext Ultra II FS DNA Library Prep with Sample Purification Beads
- #E7645, NEBNext Ultra II DNA Library Prep Kit for Illumina
- #E7103, NEBNext Ultra II DNA Library Prep with Sample Purification Beads
- #E7595, NEBNext Ultra II Ligation Module

**Please refer to the library preparation kit manual for additional required materials that are not included.**

## Index Pooling Guidelines: 96 Reaction Kit



For a link to download a sample sheet with the index sequences for use with the Illumina Experiment Manager (IEM) please go to our [FAQ's](#); or [Usage Guidelines](#) tab on [www.neb.com/E7395](http://www.neb.com/E7395) – NEBNext Multiplex Oligos for Illumina (Unique Dual Index UMI Adaptors DNA Set 1) (NEB #E7395).

For all HiSeq<sup>®</sup>/MiSeq<sup>®</sup> sequencers, Illumina uses a red laser/LED to sequence bases A and C and a green laser/LED to sequence bases G and T. For each cycle, both the red and the green channel need to be read to ensure proper image registration (i.e., A or C must be in each cycle, and G or T must be in each cycle). If this color balance is not maintained, sequencing the index read could fail. Table 1 lists some valid combinations (up to 8-plex) that can be sequenced together. For combinations > 8 choose any column and add any plex combinations as needed.

For the NovaSeq<sup>®</sup>/NextSeq<sup>®</sup>/MiniSeq<sup>®</sup> which utilize 2 color chemistry, valid index combinations must include some indices that do not start with GG in the first two cycles. Use Table 1 for some suggested combinations.

**Table 1. Index Pooling Guidelines**

PLEX	WELL POSITION
< 4	Not recommended
4	A6, B6, C6, D6 A12, B12, C12, D12 B6, C6, D6, E6 B12, C12, D12, E12 C1, D1, E1, F1 C7, D7, E7, F7 E4, F4, G4, H4 E10, F10, G10, H10
5	A1, B1, C1, D1, E1 A6, B6, C6, D6, E6 A7, B7, C7, D7, E7 A12, B12, C12, D12, E12 B1, C1, D1, E1, F1 B6, C6, D6, E6, F6 B7, C7, D7, E7, F7 B12, C12, D12, E12, F12 C1, D1, E1, F1, G1 C2, D2, E2, F2, G2 C4, D4, E4, F4, G4 C7, D7, E7, F7, G7 C8, D8, E8, F8, G8 C10, D10, E10, F10, G10 D4, E4, F4, G4, H4 D10, E10, F10, G10, H10
6-7	Any 5 plex plus 1-2 adjacent wells from the same column
8	Any column

**Table 2. Examples of “good” and “bad” index combinations based on HiSeq/MiSeq guidelines. Each index sequence is color coded to correspond to the red/green channel. For combinations of valid indices, ensure that you will have signal in both the red and green channels in each cycle.**

BAD																								
WELL POSITION	EXPECTED i7 INDEX READ								EXPECTED i5 INDEX READ															
									MiSeq, HiSeq 2000/2500					HiSeq 3000/4000, HiSeq X										
E8	T	A	T	G	G	C	A	C	T	T	G	C	G	A	G	A	T	C	T	C	G	C	A	A
F8	G	A	A	T	C	A	C	C	G	A	A	C	G	A	A	G	C	T	T	C	G	T	T	C
G8	G	T	A	A	G	G	T	G	C	G	A	A	T	T	G	C	G	C	A	A	T	T	C	G
H8	C	G	A	G	A	G	A	A	G	G	A	A	G	A	G	A	T	C	T	C	T	T	C	C
	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	X	X	✓	✓	✓	✓	✓	✓	X	X	✓	✓	✓
A1	T	T	A	C	C	G	A	C	C	G	A	A	T	A	C	G	C	G	T	A	T	T	G	G
B1	T	C	G	T	C	T	G	A	G	T	C	C	T	T	G	A	T	C	A	A	G	G	A	C
C1	T	T	C	C	A	G	G	T	C	A	G	T	G	C	T	T	A	A	G	C	A	C	T	G
D1	T	A	C	G	G	T	C	T	T	C	C	A	T	T	G	C	G	C	A	A	T	G	G	A
	X	✓	✓	✓	✓	X	✓	✓	✓	✓	✓	✓	X	✓	✓	✓	✓	✓	✓	X	✓	✓	✓	✓
GOOD																								
WELL POSITION	EXPECTED i7 INDEX READ								EXPECTED i5 INDEX READ															
									MiSeq, HiSeq 2000/2500					HiSeq 3000/4000, HiSeq X										
C1	T	T	C	C	A	G	G	T	C	A	G	T	G	C	T	T	A	A	G	C	A	C	G	G
D1	T	A	C	G	G	T	C	T	T	C	C	A	T	T	G	C	G	C	A	A	T	G	G	A
E1	A	A	G	A	C	C	G	T	G	T	C	G	A	T	T	G	C	A	A	T	C	G	A	C
F1	C	A	G	G	T	T	C	A	A	T	A	A	C	G	C	C	G	G	C	G	T	T	A	T
	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓
A12	C	G	G	C	A	T	T	A	G	T	C	A	G	T	C	A	T	G	A	C	T	G	C	C
B12	C	A	C	G	C	A	A	T	C	C	T	T	C	C	A	T	A	T	G	G	A	A	G	G
C12	G	G	A	A	T	G	T	C	A	G	G	A	A	C	A	C	G	T	G	T	T	C	C	T
D12	T	G	G	T	G	A	A	G	C	T	T	A	C	A	G	C	G	C	T	G	T	A	A	G
	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓

The index adaptor sequences for different Illumina sequencer input sheets are indicated in Table 4.

**Table 3. Examples of “good” and “bad” index combinations based on NovaSeq, NextSeq and MiniSeq guidelines. NovaSeq, NextSeq and MiniSeq use 2 color channel sequencing to simplify nucleotide detection. Clusters only in red or green are interpreted as C or T, respectively. Clusters in both red and green are read as A, while unlabeled clusters are G bases. For multiplexing a small number of samples, make sure the final index pool contains some indices that do not start with GG in the first two cycles.**

GOOD																								
WELL POSITION	EXPECTED i7 INDEX READ							EXPECTED i5 INDEX READ																
								NovaSeq					MiniSeq, NextSeq											
A12	C	G	G	C	A	T	T	A	G	T	C	A	G	T	C	A	T	G	A	C	T	G	C	C
B12	C	A	C	G	C	A	A	T	C	C	T	T	C	C	A	T	A	T	G	G	A	A	G	G
C12	G	G	A	A	T	G	T	C	A	G	G	A	A	C	A	C	G	T	G	T	T	C	C	T
D12	T	G	G	T	G	A	A	G	C	T	T	A	C	A	G	C	G	C	T	G	T	A	A	G
	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓

BAD																								
WELL POSITION	EXPECTED i7 INDEX READ							EXPECTED i5 INDEX READ																
								NovaSeq					MiniSeq, NextSeq											
C12	G	G	A	A	T	G	T	C	A	G	G	A	A	C	A	C	G	T	G	T	T	C	C	T
E12	G	G	A	C	A	T	C	A	T	A	C	C	T	G	C	A	T	G	C	A	G	G	T	A
F12	G	G	T	G	T	A	C	A	A	G	A	C	G	C	T	A	T	A	G	C	G	T	C	T
G11	G	G	T	T	G	A	A	C	T	C	C	A	C	G	T	T	A	A	C	G	T	G	G	A
	X	X	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓	✓

## Index Sequences

Table 4. Index Sequences (color coded based on HiSeq/MiSeq guidelines).

WELL POSITION	EXPECTED i7 INDEX READ	EXPECTED i5 INDEX READ	
		NovaSeq, MiSeq, HiSeq 2000/2500 (PE Flow Cell), HiSeq 3000/4000 (Single Read Flow Cell)	NextSeq, MiSeq, HiSeq 2000/2500 (Single Read Flow Cell), HiSeq 3000/4000/HiSeq X (PE Flow Cell)
A1	TTACCGAC	CGAATACG	CGTATTCTG
B1	TCGTCTGA	GTCCTTGA	TCAAGGAC
C1	TTCCAGGT	CAGTGCTT	AAGCACTG
D1	TACGGTCT	TCCATTGC	GCAATGGA
E1	AAGACCGT	GTCGATTG	CAATCGAC
F1	CAGGTTCA	ATAACGCC	GGCGTTAT
G1	TAGGAGCT	GCCTTAAC	GTTAAGGC
H1	TACTCCAG	GGTATAGG	CCTATACC
A2	AGTGACCT	TCTAGGAG	CTCCTAGA
B2	AGCCTATC	TGCGTAAC	GTTACGCA
C2	TCATCTCC	CTTGCTAG	CTAGCAAG
D2	CCAGTATC	AGCGAGAT	ATCTCGCT
E2	TTGCGAGA	TATGGCAC	GTGCCATA
F2	GAACGAAG	GAATCACC	GGTGATTC
G2	CGAATTGC	GTAAGGTG	CACCTTAC
H2	GGAAGAGA	CGAGAGAA	TTCTCTCG
A3	TCGGATTC	CGCAACTA	TAGTTGCG
B3	CTGTACCA	CACAGACT	AGTCTGTG
C3	GAGAGTAC	TGGAAGCA	TGCTTCCA
D3	TCTACGCA	CAATAGCC	GGCTATTG
E3	GCAATTCC	CTCGAACA	TGTTCGAG
F3	CTCAGAAG	GGCAAGTT	AACTTGCC
G3	GTCCTAAG	AGCTACCA	TGGTAGCT
H3	GCGTTAGA	CAGCATACT	GTATGCTG
A4	CAAGGTAC	CGTATCTC	GAGATACG
B4	AGACCTTG	TTACGTGC	GCACGTAA
C4	GTCGTTAC	AGCTAAGC	GCTTAGCT
D4	GTAACCGA	AAGACACC	GGTGTCTT
E4	GAATCCGT	CAACTCCA	TGGAGTTG
F4	CATGAGCA	GATCTTGC	GCAAGATC
G4	CTTAGGAC	CTTCACTG	CAGTGAAG
H4	ATCTGACC	CTCGACTT	AAGTCGAG
A5	TCCTCATG	GTACACCT	AGGTGTAC
B5	AGGATAGC	CCAAGGTT	AACCTTGG
C5	GGAGGAAT	GAACGGTT	AACCGTTC
D5	GACGTCAT	CCAGTTGA	TCAACTGG
E5	CCGCTTAA	GTCATCGT	ACGATGAC
F5	GACGAACT	CAATGCGA	TCGCATTG
G5	TCCACGTT	GGTTGAAC	GTTCAACC
H5	AACCGAG	CTTCGGTT	AACCGAAG

WELL POSITION	EXPECTED i7 INDEX READ	EXPECTED i5 INDEX READ	
		NovaSeq, MiSeq, HiSeq 2000/2500 (PE Flow Cell), HiSeq 3000/4000 (Single Read Flow Cell)	NextSeq, MiSeq, HiSeq 2000/2500 (Single Read Flow Cell), HiSeq 3000/4000/HiSeq X (PE Flow Cell)
A6	<b>GTCAGTCA</b>	<b>CGGCATTA</b>	<b>TAATGCCG</b>
B6	<b>CCTTCCAT</b>	<b>CACGCAAT</b>	<b>ATTGCGTG</b>
C6	<b>AGGAACAC</b>	<b>GGAATGTC</b>	<b>GACATTCC</b>
D6	<b>CTTACAGC</b>	<b>TGGTGAAG</b>	<b>CTTCACCA</b>
E6	<b>TACCTGCA</b>	<b>GGACATCA</b>	<b>TGATGTCC</b>
F6	<b>AGACGCTA</b>	<b>GGTGTACA</b>	<b>TGTACACC</b>
G6	<b>CAACACAG</b>	<b>GATAGCCA</b>	<b>TGGCTATC</b>
H6	<b>GTACCACA</b>	<b>CCACAACA</b>	<b>TGTTGTGG</b>
A7	<b>CGAATACG</b>	<b>TTACCGAC</b>	<b>GTCGGTAA</b>
B7	<b>GTCCTTGA</b>	<b>TCGTCTGA</b>	<b>TCAGACGA</b>
C7	<b>CAGTGCTT</b>	<b>TTCCAGGT</b>	<b>ACCTGGAA</b>
D7	<b>TCCATTGC</b>	<b>TACGGTCT</b>	<b>AGACCGTA</b>
E7	<b>GTCGATTG</b>	<b>AAGACCGT</b>	<b>ACGGTCTT</b>
F7	<b>ATAACGCC</b>	<b>CAGGTTCA</b>	<b>TGAACCTG</b>
G7	<b>GCCTTAAC</b>	<b>TAGGAGCT</b>	<b>AGCTCCTA</b>
H7	<b>GGTATAGG</b>	<b>TACTCCAG</b>	<b>CTGGAGTA</b>
A8	<b>TCTAGGAG</b>	<b>AGTGACCT</b>	<b>AGGTCACT</b>
B8	<b>TGCGTAAC</b>	<b>AGCCTATC</b>	<b>GATAGGCT</b>
C8	<b>CTTGCTAG</b>	<b>TCATCTCC</b>	<b>GGAGATGA</b>
D8	<b>AGCGAGAT</b>	<b>CCAGTATC</b>	<b>GATACTGG</b>
E8	<b>TATGGCAC</b>	<b>TTGCGAGA</b>	<b>TCTCGCAA</b>
F8	<b>GAATCACC</b>	<b>GAACGAAG</b>	<b>CTTCGTTC</b>
G8	<b>GTAAGGTG</b>	<b>CGAATTGC</b>	<b>GCAATTCC</b>
H8	<b>CGAGAGAA</b>	<b>GGAAGAGA</b>	<b>TCTCTTCC</b>
A9	<b>CGCAACTA</b>	<b>TCGGATTC</b>	<b>GAATCCGA</b>
B9	<b>CACAGACT</b>	<b>CTGTACCA</b>	<b>TGGTACAG</b>
C9	<b>TGGAAGCA</b>	<b>GAGAGTAC</b>	<b>GTA CTCTC</b>
D9	<b>CAATAGCC</b>	<b>TCTACGCA</b>	<b>TGCGTAGA</b>
E9	<b>CTCGAACA</b>	<b>GCAATTCC</b>	<b>GGAATTGC</b>
F9	<b>GGCAAGTT</b>	<b>CTCAGAAG</b>	<b>CTTCTGAG</b>
G9	<b>AGCTACCA</b>	<b>GTCCTAAG</b>	<b>CTTAGGAC</b>
H9	<b>CAGCATA C</b>	<b>GCGTTAGA</b>	<b>TCTAACGC</b>
A10	<b>CGTATCTC</b>	<b>CAAGGTAC</b>	<b>GTACCTTG</b>
B10	<b>TTACGTGC</b>	<b>AGACCTTG</b>	<b>CAAGGTCT</b>
C10	<b>AGCTAAGC</b>	<b>GTCGTTAC</b>	<b>GTAACGAC</b>
D10	<b>AAGACACC</b>	<b>GTAACCGA</b>	<b>TCGGTTAC</b>
E10	<b>CAACTCCA</b>	<b>GAATCCGT</b>	<b>ACGGATTC</b>
F10	<b>GATCTTGC</b>	<b>CATGAGCA</b>	<b>TGCTCATG</b>
G10	<b>CTTCACTG</b>	<b>CTTAGGAC</b>	<b>GTCCTAAG</b>
H10	<b>CTCGACTT</b>	<b>ATCTGACC</b>	<b>GGTCAGAT</b>

WELL POSITION	EXPECTED i7 INDEX READ	EXPECTED i5 INDEX READ	
		NovaSeq, MiSeq, HiSeq 2000/2500 (PE Flow Cell), HiSeq 3000/4000 (Single Read Flow Cell)	NextSeq, MiSeq, HiSeq 2000/2500 (Single Read Flow Cell), HiSeq 3000/4000/HiSeq X (PE Flow Cell)
A11	<b>GTACACCT</b>	<b>TCCTCATG</b>	<b>CATGAGGA</b>
B11	<b>CCAAGGTT</b>	<b>AGGATAGC</b>	<b>GCTATCCT</b>
C11	<b>GAACGGTT</b>	<b>GGAGGAAT</b>	<b>ATTCCTCC</b>
D11	<b>CCAGTTGA</b>	<b>GACGTCAT</b>	<b>ATGACGTC</b>
E11	<b>GTCATCGT</b>	<b>CCGCTTAA</b>	<b>TTAAGCGG</b>
F11	<b>CAATGCGA</b>	<b>GACGAACT</b>	<b>AGTTGCTC</b>
G11	<b>GGTTGAAC</b>	<b>TCCACGTT</b>	<b>AACGTGGA</b>
H11	<b>CTTCGGTT</b>	<b>AACCAGAG</b>	<b>CTCTGGTT</b>
A12	<b>CGGCATTA</b>	<b>GTCAGTCA</b>	<b>TGACTGAC</b>
B12	<b>CACGCAAT</b>	<b>CCTTCCAT</b>	<b>ATGGAAGG</b>
C12	<b>GGAAATGTC</b>	<b>AGGAACAC</b>	<b>GTGTTCTT</b>
D12	<b>TGGTGAAG</b>	<b>CTTACAGC</b>	<b>GCTGTAAG</b>
E12	<b>GGACATCA</b>	<b>TACCTGCA</b>	<b>TGCAGGTA</b>
F12	<b>GGTGTACA</b>	<b>AGACGCTA</b>	<b>TAGCGTCT</b>
G12	<b>GATAGCCA</b>	<b>CAACACAG</b>	<b>CTGTGTTG</b>
H12	<b>CCACAACA</b>	<b>GTACCACA</b>	<b>TGTGGTAC</b>

## Sequencing on the Illumina Platform

Pool equal molar amounts of libraries for sequencing on the Illumina platforms using the cycle settings shown in the table below.

RUN SEGMENT	CYCLE NUMBER
Read 1	X defined by users
Index 1 (i7)	8 (without UMI)
	19 (with UMI)
Index 2 (i5)	8
Read 2	X defined by users

## Kit Components

The NEBNext Multiplex Oligos for Illumina (Unique Dual Index UMI Adaptors DNA Set 1) are functionally validated through library preparation using the NEBNext Library Prep Kits and sequencing on the Illumina platforms.

### NEB #E7395S Table of Components

NEB #	CONCENTRATION	PRODUCT	VOLUME
E7396A	20 µM	NEBNext UMI DNA Adaptor Plate	1 plate (2.5 µl/well)
E7397A	40 µM (Total)	NEBNext Primer Mix	0.48 ml
E7398A		NEBNext UMI Adaptor Dilution Buffer	5 ml

### NEB #E7395L Table of Components

NEB #	CONCENTRATION	PRODUCT	VOLUME
E7396A	20 µM	NEBNext UMI DNA Adaptor Plate	4 plates (2.5 µl/well)
E7397AA	40 µM (Total)	NEBNext Primer Mix	2 x 0.96 ml
E7398AA		NEBNext UMI Adaptor Dilution Buffer	20 ml

## Revision History

REVISION #	DESCRIPTION	DATE
1.0	N/A	3/20

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