Abundance of fungal hyphae din seawater by epifluorescense microscopy using Calcofluor White stain method

Prepared by M.H. Gutiérrez, May 2018

- To collect samples in sterile 50 mL polypropylene tubes and fix with formaldehyde or glutaraldehyde (2% final concentration). Store samples at 4 °C in the dark.
- To filter 5 to 30 mL (depends on the environment) of seawater by 0.22-µm mesh black 25 mm diameter polycarbonate filters (Millipore Corp.).
- To stain filters with the retained material directly with 600 μ L of aqueous 0.1% Calcofluor White, making sure to cover the entire area of the filter. After a few minutes (5-10 m) use vacuum to remove the excess of stain from the filter. Avoid any light source.
- Place the filter (sample side up) onto slide and add 1 drop of non-fluorescent immersion oil on top of the filter and cover with a cover slip.
- The slides can be counted immediately on the epifluorescent microscope or stored frozen (-20 °C).
- In a epifluorescence microscope equipped with UV filter (used for DAPI, eg. filter set 49 Carl Zeiss Ltd., 365 nm excitation and 445-450 nm emission band pass) to examine at 1000X the entire effective area of the filters.
- To count all hyphae identified and record their length and width, use cylinder volume as a morphological approximation to estimate the biovolume of fungal filaments. Biomass can be estimated based from biomass:biovolume ratios described fro fungi (Newell and Statzell-Tallman, 1982).

References

Damare S, Raghukumar C (2008) Fungi and macroaggregation in deep-sea sediments. Microb Ecol 56:168–177

Jebarag CS, Raghukumar C (2009) Anaerobic denitrification in fungi from the coastal marine sediments off Goa, India. Mycol Res 113:100-109

Newell, S.Y., Statzell-Tallman, A. (1982) Factors for conversion of fungal biovolume values to biomass, carbon and nitrogen: variation with mycelial ages, growth conditions, and strains of fungi from a salt marsh. *Oikos* **39**: 261-268

Rasconi S, Jobard M, Jouve L, Sime-Ngando T (2009) Use of calcofluor white for detection, identification, and quantification of phytoplanktonic fungal parasites. Appl Environ Microbiol 75:2545-2553