Cell culture, transfection, and imaging

Authors: Will Hancock-Cerutti^{1,2,3}, Pietro De Camilli^{1,3}

¹Departments of Neuroscience and of Cell Biology, Howard Hughes Medical Institute, Program in Cellular Neuroscience, Neurodegeneration and Repair, Yale University School of Medicine, New Haven, Connecticut 06510, USA

²Interdisciplinary Neuroscience Program and MD-PhD Program, Yale University School of Medicine, New Haven, Connecticut 06510, USA

³Aligning Science Across Parkinson's (ASAP) Collaborative Research Network, Chevy Chase, MD, 20815

Abstract

This protocol describes general procedures for culturing HeLa cells, transient transfection, and imaging using an Andor Dragonfly spinning disk confocal system.

Keywords

Cell culture, transfection, spinning-disk confocal, fluorescent microscopy.

Solutions to prepare

DMEM containing 10% FBS, 100 U/ml penicillin, 100 mg/ml streptomycin, and 2 mM L-glutamine (all from Gibco).

A. General preparation

- HeLa-M cells were cultured at 37°C in 5% CO2 and DMEM containing 10% FBS, 100 U/ml penicillin, 100 mg/ml streptomycin, and 2 mM L-glutamine (all from Gibco).
 Note: For general maintenance, when cells reached 80-90% confluency, they were deattached from the dish with Trypsin and diluted 1:20 in a new dish.
- For live-cell imaging experiments, cells were seeded on glass-bottomed dishes (MatTek) at a concentration of 35,000 cells per dish and transfected after 24h using FuGene HD (Promega) in Opti-MEM (Gibco).
- 3. Cell were imaged 24 hours after transfection
- 4. Just before imaging, the growth medium was removed and replaced with prewarmed live-cell imaging solution (Life Technologies).
- 5. For lysotracker experiments, cells were incubated in 50 nM LysoTracker Red DND-99 (ThermoFisher) in complete DMEM for 30 minutes, washed twice with media, then imaged in live-cell imaging solution.
- 6. All live-cell imaging was performed at 37°C and 5% CO₂.
- 7. Spinning-disk confocal microscopy was performed using an Andor Dragonfly system equipped with a plan apochromat objective (63×, 1.4 NA, oil) and a Zyla scientific CMOS camera.
- 8. For any given experiment, the same exposure time, laser power, and gain were used for image acquisition to allow for quantitative comparison.

B. Imaging of cells stably expressing STING-GFP

- 1. Cells stably expressing STING-GFP were generated as described elsewhere.
- 2. Stable STING-GFP HeLa-M cells were cultured at 37°C in 5% CO₂ and DMEM containing 10% FBS, 100 U/ml penicillin, 100 mg/ml streptomycin, and 2 mM L-glutamine (all from Gibco).
- For experiments using siRNA, 60 pmols of the indicated siRNA was transfected using 6 μL Lipofectamine RNAiMax (ThermoFisher) in Opti-MEM (Gibco) per dish according to manufacturer protocol. Cells were imaged 72 hours after siRNA transfection.
- For experiments using cGAMP, 50 μg/L of cGAMP was transfected using using 18 μL Lipofectamine RNAiMax (ThermoFisher) in Opti-MEM (Gibco) per dish according to manufacturer protocol. Cells were imaged 14 hours after transfection.