**Light microscopy immunoperoxidase staining protocol**

1. Select sections to be processed from the brain tissue bank.
2. Wash sections thoroughly with a phosphate-buffered saline (PBS, 0.01M, pH 7.4) solution.
3. Treat the sections at room temperature with a 1% sodium borohydride (NaBH4) solution in PBS for 20 minutes.
4. Rinse sections thoroughly in PBS
5. Pre-incubate sections for 1h at room temperature in a solution containing 1% normal serum (from the species used to generate the secondary antibodies), 0.3% Triton X-100, and 1% bovine serum albumin (BSA) in PBS.
6. Incubate the sections for 24 h at room temperature in a solution containing the primary antibodies in 1% normal serum, 0.3% Triton X-100, and 1% BSA in PBS.
7. Next day, thoroughly rinse the sections in PBS
8. Incubate the sections in a PBS solution containing the appropriate biotinylated secondary antibody (1:200; Vector Labs, Burlingame, CA, USA) combined with 1% normal serum, 0.3% Triton X-100, and 1% BSA for 90 minutes at room temperature.
9. Wash the sections in PBS
10. Incubate the sections in an avidin-biotin-peroxidase complex (ABC; 1:100; Vector Labs, Burlingame, CA, USA) solution for 90 minutes at room temperature.
11. Rinse the sections in PBS twice followed by a third rinse in TRIS buffer (0.05M; pH 7.6).
12. Incubate the sections in a solution containing 0.025% 3,3ʹ-diaminobenzidine tetrahydrochloride, 10 mM imidazole, and 0.005% hydrogen peroxide in Tris buffer for 10 minutes at room temperature.
13. Rinse the sections with PBS
14. Mount sections onto gelatin-coated slides, and coverslip with Permount.
15. Digitalize the slides with an Aperio ScanScope CS system (Aperio Technologies)