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Reporting Summary

Nature Research wishes to improve the reproducibility of the work that we publish. This form provides structure for consistency and transparency in reporting. For further information on Nature Research policies, see our Editorial Policies and the Editorial Policy Checklist.

Statistics

For all statistical analyses, confirm that the following items are present in the figure legend, table legend, main text, or Methods section.					
n/a	Cor	Confirmed			
\boxtimes		The exact sample size (n) for each experimental group/condition, given as a discrete number and unit of measurement			
\boxtimes		A statement on whether measurements were taken from distinct samples or whether the same sample was measured repeatedly			
\boxtimes		The statistical test(s) used AND whether they are one- or two-sided Only common tests should be described solely by name; describe more complex techniques in the Methods section.			
\boxtimes		A description of all covariates tested			
\boxtimes		A description of any assumptions or corrections, such as tests of normality and adjustment for multiple comparisons			
\boxtimes		A full description of the statistical parameters including central tendency (e.g. means) or other basic estimates (e.g. regression coefficient) AND variation (e.g. standard deviation) or associated estimates of uncertainty (e.g. confidence intervals)			
\boxtimes		For null hypothesis testing, the test statistic (e.g. F, t, r) with confidence intervals, effect sizes, degrees of freedom and P value noted Give P values as exact values whenever suitable.			
\boxtimes		For Bayesian analysis, information on the choice of priors and Markov chain Monte Carlo settings			
\boxtimes		For hierarchical and complex designs, identification of the appropriate level for tests and full reporting of outcomes			
\boxtimes		Estimates of effect sizes (e.g. Cohen's d, Pearson's r), indicating how they were calculated			
		Our web collection on statistics for biologists contains articles on many of the points above.			

Software and code

Policy information about <u>availability of computer code</u>						
Data collection	For image aquisition: Harmony High-Content Imaging and Analysis Software (version 4.7) For probe set design: Oligostan-HT (https://hub.docker.com/r/oligostan/oligostan_ht_rna) For image visualization: Image J 1.53d					
Data analysis	The homemade code used to design oligo pool probe setsis found at https://hub.docker.com/r/oligostan/oligostan_ht_rna FISH-quant and FISH-quant v2 for image analysis are available at https://github.com/fish-quant					

For manuscripts utilizing custom algorithms or software that are central to the research but not yet described in published literature, software must be made available to editors and reviewers. We strongly encourage code deposition in a community repository (e.g. GitHub). See the Nature Research guidelines for submitting code & software for further information.

Data

Policy information about availability of data

All manuscripts must include a data availability statement. This statement should provide the following information, where applicable:

- Accession codes, unique identifiers, or web links for publicly available datasets
- A list of figures that have associated raw data

- A description of any restrictions on data availability

An example of an oligo pool is provided as a Supplementary Table

Field-specific reporting

Life sciences

Please select the one below that is the best fit for your research. If you are not sure, read the appropriate sections before making your selection.

Behavioural & social sciences Ecological, evolutionary & environmental sciences

For a reference copy of the document with all sections, see <u>nature.com/documents/nr-reporting-summary-flat.pdf</u>

Life sciences study design

All studies must disclose on these points even when the disclosure is negative.

Sample size	NA
Data exclusions	NA
Replication	NA
Randomization	NA
Blinding	NA

Reporting for specific materials, systems and methods

Methods

We require information from authors about some types of materials, experimental systems and methods used in many studies. Here, indicate whether each material, system or method listed is relevant to your study. If you are not sure if a list item applies to your research, read the appropriate section before selecting a response.

Materials & experimental systems

n/a	Involved in the study	n/a	Involved in the study
\boxtimes	Antibodies	\boxtimes	ChIP-seq
	Eukaryotic cell lines	\boxtimes	Flow cytometry
\boxtimes	Palaeontology and archaeology	\boxtimes	MRI-based neuroimaging
	Animals and other organisms		
\boxtimes	Human research participants		
\boxtimes	Clinical data		
\square	Dual use research of concern		

Eukaryotic cell lines

Policy information about <u>cell lines</u>	
Cell line source(s)	HeLa Kyoto cells were a gift from the lab of A. Hyman and used in Poser, I., Sarov, M., Hutchins, J. et al. BAC TransgeneOmics: a high-throughput method for exploration of protein function in mammals. Nat Methods 5, 409–415 (2008). https:// doi.org/10.1038/nmeth.1199
	HeLa cells expressing centrin 1-GFP were a gift from the lab of B. Delaval and originated from: Matthieu Piel, Pablo Meyer, Alexey Khodjakov, Conly L. Rieder, Michel Bornens; The Respective Contributions of the Mother and Daughter Centrioles to Centrosome Activity and Behavior in Vertebrate Cells. J Cell Biol 17 April 2000; 149 (2): 317–330. doi: https:// doi.org/10.1083/jcb.149.2.317
	HEK293T cells stably expressing GFP-LSM14A were a gift from the lab of D. Weil and originated from: Arnaud Hubstenberger, Maïté Courel, Marianne Bénard, et al.P-Body Purification Reveals the Condensation of Repressed mRNA Regulons, Molecular Cell,Volume 68, Issue 1,2017,Pages 144-157.e5,ISSN 1097-2765, https://doi.org/10.1016/j.molcel.2017.09.003.
Authentication	All cell lines showed GFP fluorescence reflecting the tagged protein's localization
Mycoplasma contamination	Cell lines were tested negative for Mycoplasma
Commonly misidentified lines (See <u>ICLAC</u> register)	No commonly misidentified cell lines were used in this study

Animals and other organisms

Policy information about studies involving animals; ARRIVE guidelines recommended for reporting animal research				
Laboratory animals	Male Black-6 Mice (C57BL/6), 6 weeks old at time of sacrifice were used. Mice were housed in temperature-controlled ventilated cages (20–22 °C) with a 12 h light-dark cycle, with percentage of humidity between 45 and 55%, and maintained in pathogen-free conditions in the institute's animal facility.			
Wild animals	The study did not involve wild animals.			
Field-collected samples	The study did not involve samples colleted from the field.			
Ethics oversight	Mouse experiments were performed in strict accordance with the guidelines of the European Community (86/609/EEC) and the French National Committee (87/848) for care and use of laboratory animals. They comply with the ARRIVE guidelines and were approved by the French Ministry of Higher Education, Research and Innovation (reference APAFIS#18685) to be performed in the institute animal facility (agreement # F3417216).			

Note that full information on the approval of the study protocol must also be provided in the manuscript.